

A NEW ANTI-BIOFILM HYDROFIBER® DRESSING: IN VITRO DETERMINATION OF MICROBIAL KILL RATE IN BIOFILMS

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Introduction

In their natural habitat, bacteria usually exist in communities encapsulated in a matrix of gelatinous extracellular polymeric substances (EPS) which is attached to a surface. This is known as biofilm.¹ Biofilm EPS is produced by the bacteria and serves as a protective niche for survival. It renders them more tolerant to the host immune response and antimicrobial therapies alike.^{1,2} Biofilm is increasingly recognised as being a barrier to wound healing and there is a growing evidence that biofilms are a cause of wound chronicity^{1,2,3} – with studies demonstrating biofilm prevalence of at least 60% in chronic wounds.³ Therefore, to be clinically useful, antimicrobial therapies must be effective against biofilm.

Aim

This poster describes *in-vitro* studies used to measure the anti-biofilm properties of a new, custom-designed anti-biofilm Hydrofiber® dressing, AQUACEL® Ag+.

Method

Study 1

Biofilms of challenge organisms (Fig 1) were grown on sterile cotton gauzes by immersion in a bacteria-containing simulated wound fluid at 35°C for ≥ 48 hours. Gauzes were then rinsed with isotonic saline to remove any unattached bacteria. The presence of biofilm was confirmed by scanning electron microscopy (SEM). Biofilm-colonised gauzes were exposed to test dressings (Fig 2) hydrated with isotonic saline for 4, 24 and 48 hours at 35°C. Bacterial bioburden was determined by processing the gauzes in a neutralising diluent and then assaying by standard microbiological techniques.

Study 2

Biofilms were prepared as in study 1, then a PHMB gauze (Fig 2) was applied for a further 48 hours to eliminate any planktonic cells whilst maintaining growth of a predominately biofilm population. AQUACEL® Ag+ dressing samples were then applied hydrated with simulated wound fluid and incubated at 35°C for up to 96 hours. Quantitative analysis was performed as previously described.

Figure 1. Challenge organisms

Study	Challenge organism
1	<i>Pseudomonas aeruginosa</i> (NCIMB 8626)
1	<i>Staphylococcus aureus</i> (NCIMB 9518)
1	<i>Candida albicans</i> (NCPF 3179)
2	<i>P. aeruginosa</i> (wild type strain PA01)

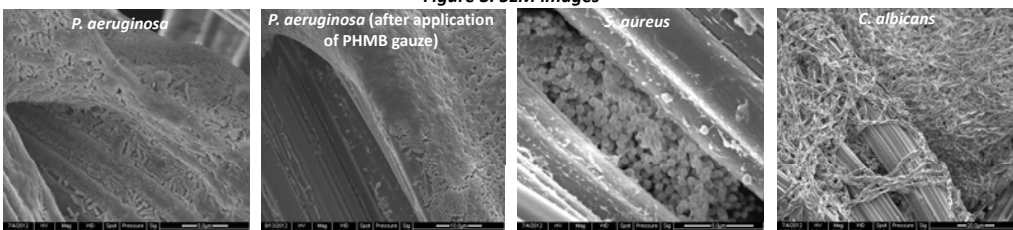
Figure 2. Test dressings

Coding	Material	Product	Manufacturer
ABHF	Anti-biofilm Hydrofiber®	AQUACEL® Ag+ (CE Marked 2013)	ConvaTec
SHF	Silver-containing Hydrofiber®	AQUACEL® Ag	ConvaTec
HF	Hydrofiber®	AQUACEL®	ConvaTec
PHMB-Gauze	Polyhexamethylene biguanide (PHMB)-containing non-adherent gauze	Telfa™ AMD	Covidien

Results

SEM confirmed the presence of biofilm on the cotton gauze substrates, including after PHMB gauze (as in Study 2; Figure 3):

Figure 3. SEM images



Results (cont.)

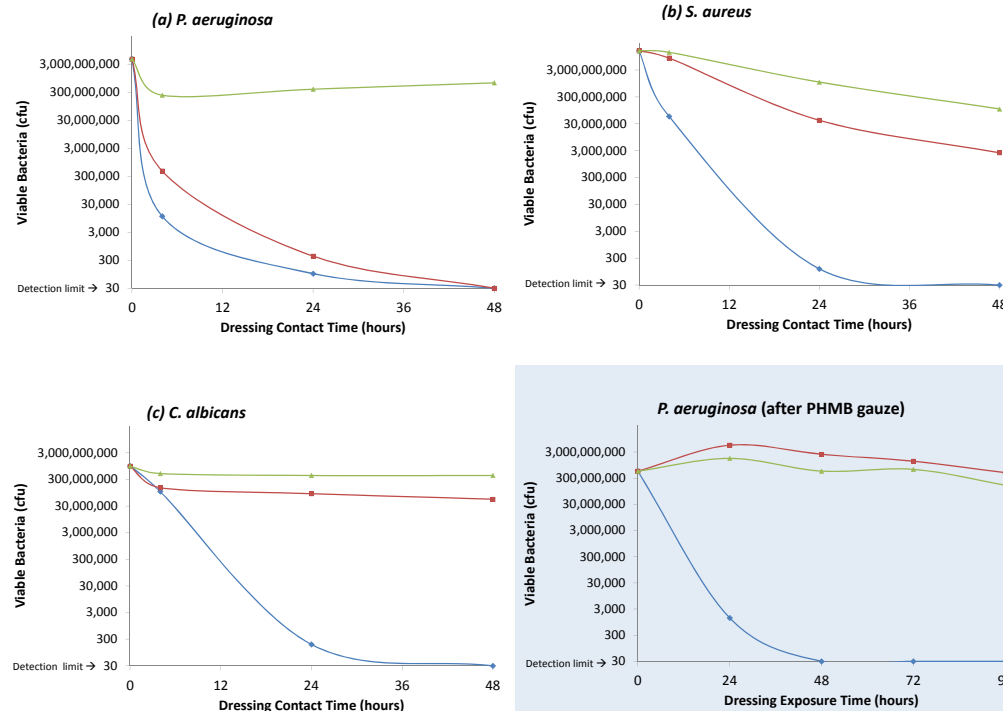


Figure 4. Study 1 – Biofilm reduction (a) *P. aeruginosa*; (b) *S. aureus*; (c) *C. albicans*. (◆) AQUACEL® Ag+; (■) AQUACEL® Ag; (▲) AQUACEL®

Figure 5. Study 2 – Biofilm reduction after PHMB gauze. (◆) AQUACEL® Ag+; (■) AQUACEL® Ag; (▲) AQUACEL®

Discussion & Conclusion

Biofilm bacteria counts of potential wound pathogens fell below the limit of detection within 48 hours for the AQUACEL® Ag+ dressing. Approximately an 8 log reduction (100 million-fold) reduction in biofilm bacteria was achieved for each challenge organism for the AQUACEL® Ag+ dressing. A >3 log reduction was achieved within 4 hours for *S. aureus* and *P. aeruginosa* biofilm bacteria.

Three potential wound pathogens were shown to form dense biofilm on cotton gauze (*S. aureus*, *P. aeruginosa* and *C. albicans*). Biofilm forms of each potential pathogen were tolerant to two antimicrobial dressings (ionic silver and PHMB). Biofilm forms of each potential pathogen were highly-susceptible to a new anti-biofilm Hydrofiber® dressing, AQUACEL® Ag+.

References:

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